

NEUTRAL OXYGEN-CONTAINING VOLATILE CONSTITUENTS OF GREEK TOBACCO*

B. KIMLAND, R. A. APPLETON, A. J. AASEN, J. ROERADE and C. R. ENZELL†

Chemical Research Department, Swedish Tobacco Co. S-10462 Stockholm, Sweden

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Abstract—The volatile fraction of the neutral, oxygen containing constituents of sun-cured Greek tobacco has been studied. Thirty-two compounds, mainly ketones and aldehydes, were identified using combined gas chromatography—mass spectrometry. The majority of these have not previously been reported to occur in tobacco and two are new natural products. Possible origins of some of the compounds encountered are discussed.

INTRODUCTION

EARLIER publications in this series on sun-cured Greek tobacco have dealt with the saturated¹ and unsaturated volatile² hydrocarbons. The present work, representing a continuation of these studies, concerns the more polar compounds in the volatile, neutral fraction which represents an important part of the flavour of this tobacco. An authoritative account of the constituents of tobacco and smoke has been given by Stedman³.

RESULTS AND DISCUSSION

The volatile neutral material of tobacco grown in Serres, Greece, in 1968 was isolated as detailed in the Experimental, whilst taking extreme precautions to avoid contamination during isolation. It was shown by GLC to be very complex and was divided by means of Girard T reagent into a carbonyl fraction, designated A, and a remainder, which in turn was separated into a hydrocarbon fraction, discussed previously,^{1,2} and a non-hydrocarbon fraction B‡. Subsequent chromatography on silica gel of the fractions containing the oxygenated constituents furnished fractions A1–A7 and B1–B8, which were studied by GLC and GLC–MS.

The identification of the compounds in Table 1 was accomplished by comparison of mass spectra and retention times with those of authentic material or,^{4–7} when the con-

* Part VI in the series "Tobacco Chemistry". For Part V see Ref. 58.

† To whom correspondence should be addressed.

‡ The use of Girard T reagent was unsatisfactory since even the separation of sterically unhindered carbonyls was far from quantitative and this procedure will hence be omitted in future studies.

¹ C. R. ENZELL, A. ROSENGREN and I. WAHLBERG, *Tobacco Sci.* **13**, 127 (1969).

² R. A. APPLETON, C. R. ENZELL and B. KIMLAND, *Beitr. Tabakforsch.* **5**, 266 (1970).

³ R. L. STEDMAN, *Chem. Revs.* **68**, 153 (1968).

⁴ *M S D C Series Mass Spectral Data Nos. 1 to 3000*, Mass Spectrometry Data Centre, Aldermaston, England.

⁵ *Dow Uncertified Mass Spectral Data* (edited by R. S. GOHLKE), Framington, Mass. (1963).

⁶ E. VON SYDOW, K. ANJOU and G. KARLSSON, *Arch. Mass Spectral Data* **1**, 385 (1970).

⁷ A. CORNU and R. MASSOT, *Compilation of Mass Spectral Data*, Heyden, London (1966).

TABLE 1 COMPONENTS IDENTIFIED IN FRACTIONS A AND B OF GREEK TOBACCO

No	Compound	Content in tobacco (ppm)	Fraction A	Fraction B	Method of identification	Ref	Prev detected S = smoke T = tobacco	Ref
Occurring in fraction A only								
1	β -Cyclocitral	0.1	1,2		MS, GC	4		
2	Benzaldehyde	0.07	2		MS, GC	5	S, T	25, 26
3	Safranal	0.1	2		MS, GC	6		
4	6-Methyl-3,5-heptadien-2-one	0.3	3		MS, GC			
5	Carvenone	0.07	3		MS, GC	6		
6	cis- ψ -Ionone	0.09	3		MS, GC	8		
7	2-Acetylpyrrole	0.3	6, 7		MS, GC		T	27
Occurring in fraction B only								
8	Pentadecanal	0.04		1	MS, GC			
9	Methyl myristate	0.2		1	MS, GC	9	T	28
10	Methyl palmitate	0.6		1	MS, GC	9	T	28
11	Ethyl palmitate	0.05		1	MS, GC	10		
12	Hexahydrofarnesylacetone	3.2		1, 2	MS, GC, NMR, IR	11, 12	S, T	11, 29
13	2,3,6-Trimethyl-1,4-naphthoquinone	0.5		1, 2	MS, GC		S	30
14	2,3-Dimethyl-1,4-naphthoquinone	0.03		2	MS, GC	13		
15	Benzyl acetate	0.4		2, 3	MS, GC	5	S, T	28, 31, 32
16	β -Phenylethyl acetate	0.6		2, 3, 4	MS, GC, NMR, IR	7	S, T	28, 32
17	Camphor	0.01		3	MS, GC	14		
18	Veratrole	0.03		4	MS, GC	15		
19	Di-n-butyl phthalate	0.04		4	MS, GC	5	T	33
20	Piperitone	0.4		5	MS, GC	14		
21	2,3-Dimethyl-4-hydroxy-2-nonenic acid lactone	0.4		6	MS, GC	16	T	16
22	Benzyl alcohol	13		7, 8	MS, GC, NMR, IR	5	S, T	28, 34
23	β -Phenylethanol	21		7, 8	MS, GC, NMR, IR	5	S, T	28, 34
Occurring in fraction A and B								
24	Nonanal	0.9	1, 2	1	MS, GC	5		
25	Decanal	0.5	1	1	MS, GC	5		
26	Damascenone	0.1	1	1	MS, GC	17	T	40
27	Farnesylacetone	0.4	2	2	MS, GC, NMR, IR	62	S	62
28	6-Methyl-2-heptanone	0.3	3	2, 3	MS, GC	7		
29	Solanone	7	1, 2, 3	2, 3, 4	MS, GC, NMR, IR, UV	11, 18	S, T	11, 18, 35, 36
30	Geranylacetone	6	3	2, 3, 4, 5	MS, GC, NMR, IR	19, 20		
31	6-Methyl-5-hepten-2-one	1	3, 5	3, 4	MS, GC	8		
32	Dihydroactinidiolide	13	6	6, 7, 8	MS, GC, NMR, IR	21-24	T	21

⁸ A F THOMAS, B WILLHALM and R MULLER, *Org Mass Spectrom* **2**, 223 (1969)

⁹ R RYHAGE and E STENHAGEN, *Arkiv Kemi* **13**, 523 (1959)

¹⁰ R RYHAGE and E STENHAGEN, *Arkiv Kemi* **14**, 483 (1959)

¹¹ L C COOK and A RODGMAN, *Tobacco Sci* **9**, 137 (1965)

¹² M STOLL, M WINTER, F GAUTSCHI, I FLAMENT and B WILLHALM, *Helv Chim Acta* **50**, 628 (1967)

¹³ S J DI MARI, J H SUPPLE and H RAPOPORT, *J Am Chem Soc* **88**, 1226 (1966)

¹⁴ E VON SYDOW, *Acta Chem Scand* **18**, 1099 (1964)

¹⁵ C S BARNES and J L OCCOLOWITZ, *Austral J Chem* **16**, 219 (1963)

¹⁶ H KANEKO and M MITA, *Agric Biol Chem* **33**, 1525 (1969)

¹⁷ E DEMOLE, P ENGIST, U SAUBERLI, M STOLL and E KOVATS, *sz, Helv Chim Acta* **53**, 541 (1970).

¹⁸ R R JOHNSON and J A NICHOLSON, *J Org Chem* **30**, 2918 (1965)

¹⁹ R G BUTTERY and R M SEIFERT, *J Agric Food Chem* **16**, 1053 (1968)

²⁰ G POPJAK, in *Natural Substances Formed Biologically from Mevalonic Acid* (edited by T W GOODWIN), p 30, Academic Press, New York (1970)

²¹ W C BAILEY, JR, A K BOSE, R M IKEDA, R H NEWMAN, H Y SECOR and C VARSEL, *J Org Chem* **33**, 2819 (1968)

²² P H CHEN, W F KUHN, F WILL and R M IKEDA, *Org Mass Spectrom* **3**, 199 (1970)

²³ S FUKUSHIMA, Y AKAHORI and T TSUNEA, *J Pharm Soc Japan* **89**, 1729 (1969)

²⁴ T SAKAN, S ISOE and S B HYEON, *Tetrahedron Letters* 1623 (1967)

²⁵ J ONISHI and M NAGASAWA, *Bull Agric Chem Soc Japan* **19**, 143 (1955)

²⁶ C NEUBERG and J BURKARD, *Biochem Z* **243**, 472 (1931)

²⁷ I ONISHI, H TOMITA and T FUKUZUMI, *Bull Agr Chem Soc Japan* **20**, 61 (1956)

stituents were present in sufficient amounts, by isolation and further spectroscopic studies. The abundances of the individual constituents (Table 1) were evaluated by integration of the peaks in the gas chromatograms, these figures are very approximate due to differences in detector response, imprecise integration and the presence of residual solvents. Moreover, some of them might be erroneous due to partial loss of the most volatile compounds and incomplete transfer of the least volatile constituents during distillation.

As shown in Table 1, nineteen of the compounds listed have not previously been detected in tobacco. All of the new compounds, with the exception of veratrole and ethyl palmitate, are either ketones or aldehydes, while all alcohols, lactones and remaining esters identified are previously known constituents of tobacco. Of the compounds belonging to the latter group, benzyl alcohol, β -phenylethanol and the corresponding acetates have also been found to occur in tobacco smoke.^{28,31,32,34}

Of the six aldehydes encountered, benzaldehyde is the only one previously found in tobacco and also in smoke.^{25,26} Of the others, β -cyclocitral and safranal are isoprenoid, while three are saturated straight chain compounds having 9, 10 and 15 carbon atoms respectively. Although all of these compounds, known to possess characteristic odors, have been encountered in Nature before, only nonanal and decanal seem to be of general occurrence.³⁷

Of the ketones three are non-isoprenoid (7,13,14) and of these only 2-acetylpyrrole has previously been found in tobacco,²⁷ while 2,3,6-trimethyl-1,4-naphthoquinone has been found in smoke.³⁰ The former compound, which also occurs in black tea,³⁸ cacao and tinctures of valerian root,²⁷ is regarded as partly responsible for the improvement of the tobacco flavour realised on adding valerian root tinctures.³⁹

Although several of the isoprenoid ketones encountered are classical perfume constituents present in many essential oils and various materials for human consumption, only solanone^{11,18,35,36} and hexahydrofarnesylacetone^{11,29} have previously been reported present in tobacco. However, Demole and Berthet⁴⁰ have kindly informed us that they have independently established the presence in tobacco of one of these isoprenoid ketones, damascenone. This is an important aroma constituent of Bulgarian rose oil (*Rosa damascena* Mill.) and its structure has recently been determined.¹⁷ Of the compounds under consideration, only solanone,^{11,18,35,36} farnesylacetone⁶² and hexahydrofarnesylacetone^{11,29} have so far been detected in smoke, but it seems highly probable that virtually all of them can be transferred to the smoke and contribute to the flavour of both tobacco and smoke. Hence they represent potential additives for improving or altering the aroma. In fact, addition of 6-methyl-5-hepten-2-one and geranylacetone, the presence of which in tobacco has been

²⁸ I. ONISHI, H. TOMITA and T. FUKUZUMI, *Bull. Agric. Chem. Soc. Japan* **21**, 239 (1957).

²⁹ J. N. SCHUMACHER and L. L. VESTAL, unpublished results cited in ref. 11.

³⁰ W. J. CHAMBERLAIN and R. L. STEDMAN, *Phytochem.* **7**, 1201 (1968).

³¹ D. BURDICK, I. SCHMELTZ, R. L. MILLER and R. L. STEDMAN, *Tob. Sci.* **7**, 97 (1963).

³² D. BURDICK and R. L. STEDMAN, *Tob. Sci.* **7**, 113 (1963).

³³ R. L. STEDMAN and M. DYMICKY, *Tob. Sci.* **3**, 57 (1959).

³⁴ M. IZAWA, Y. KOBASHI and S. SAKAGUCHI, *Bull. Agric. Chem. Soc. Japan* **21**, 364 (1957).

³⁵ S. OSMAN and J. BARSON, *Chem. & Ind.* 699 (1966).

³⁶ T. FUKUZUMI, H. KANEKO, H. TAKAHARA, H. NAKAMURA and J. ONISHI, *Nippon Senbai Kosha Chuo Kenkyusho Kenkyu Hokoku*, No. 107, 269 (1965), *Chem. Abs.* **64**, 11567c (1966).

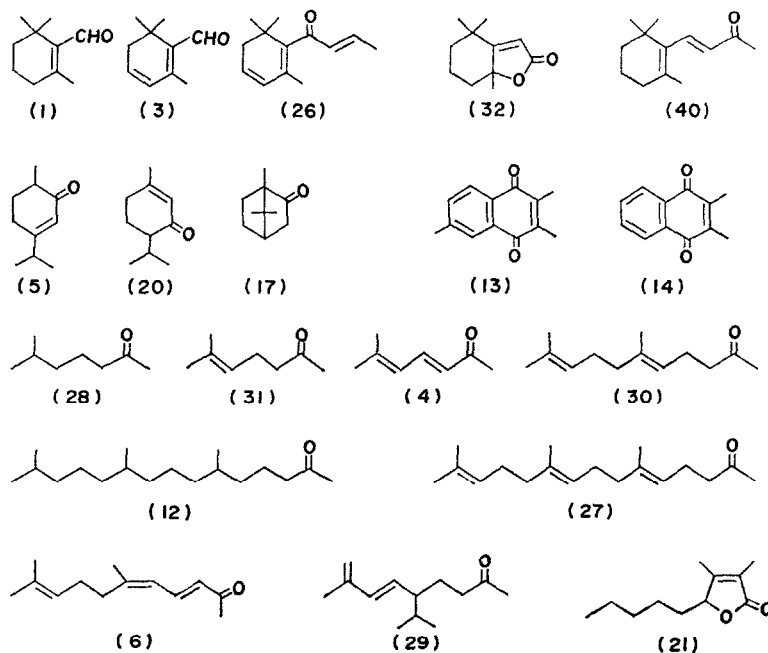
³⁷ S. ARCTANDER, *Perfume and Flavor Chemicals*, S. Arctander, Montclair, New Jersey (1969).

³⁸ R. YAMAMOTO, K. ITO and H. CHIEN, *J. Agric. Chem. Soc. Japan* **16**, 800 (1940).

³⁹ M. NIO, in *Tobacco Industry*, p. 770, Sangyo-hyoronsha, Japan (1950).

⁴⁰ E. DEMOLE and D. BERTHET, *Helv. Chim. Acta* **54**, 681 (1971).

suggested but not proved,^{41,42} and of solanone has been stated to give organoleptically improved tobacco.⁴¹ Two of the constituents, 4 and 14, are new natural products.



Several of the compounds encountered in the present study are nonisoprenoids and a large number of aromatic hydrocarbons have previously been shown to occur in the same tobacco.² It seems probable therefore that many tobacco constituents are the result of oxidative changes, which may be non-enzymatic and could occur during drying, curing and storage of the harvested leaf or even before harvest. Previous studies of tobacco diterpenoids⁴³⁻⁵¹ seem to support this, as a number of these compounds could be viewed as the result of a set of oxidation and/or rearrangement reactions, see Scheme 1 (Solid line arrows indicate that the reactions have been accomplished chemically). Thus 4,8,13-divatriene-1,3-diol (34, two diastereomers)⁴³ probably gives rise to the ketoacid (35, two diastereomers),⁴⁶ the aldehyde (36)⁴⁷ and, either directly or via the 1,5-diol (37, two diastereomers)⁴⁴ to the 5,8-oxides (38 and 39, two diastereomers).⁴⁵ The aldehyde (36) and the ketoacid (35) constitute in turn adequate precursors for solanone (29)¹⁸ satisfactorily explaining, as

⁴¹ R. B. GRIFFITH, R. R. JOHNSON and A. D. QUINN, *U.S. Pat.* 3 174 485, *Chem. Abs.* **62**, 150842e (1965).

⁴² I. W. HUGHES, private communication.

⁴³ D. L. ROBERTS and R. L. ROWLAND, *J. Org. Chem.* **27**, 3989 (1962).

⁴⁴ R. L. ROWLAND and D. L. ROBERTS, *J. Org. Chem.* **28**, 1165 (1963).

⁴⁵ R. L. ROWLAND, A. RODGMAN, J. N. SCHUMACHER, D. L. ROBERTS, L. C. COOK and W. E. WALKER, JR., *J. Org. Chem.* **29**, 16 (1964).

⁴⁶ G. W. KINZER, T. F. PAGE, JR. and R. R. JOHNSON, *J. Org. Chem.* **31**, 1797 (1966).

⁴⁷ J. L. COURTNEY and S. McDONALD, *Tetrahedron Letters* 459 (1967).

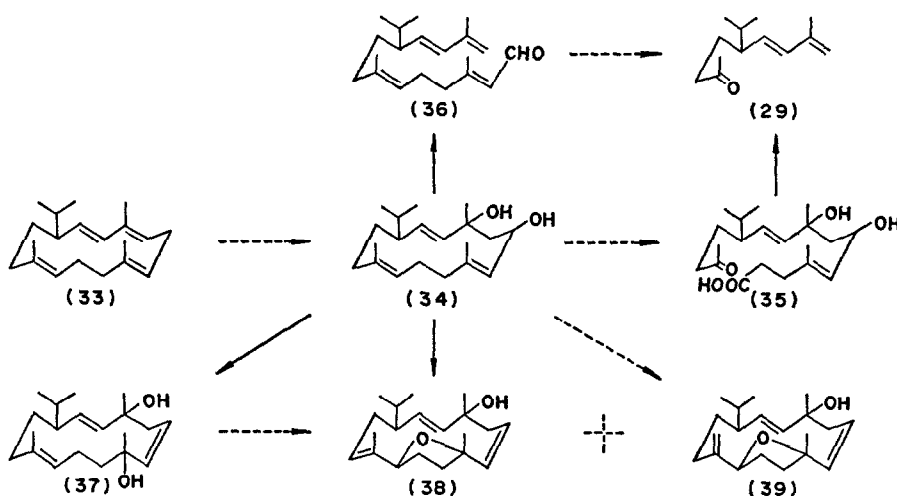
⁴⁸ W. G. DAUBEN, W. E. THIessen and P. R. RESNICK, *J. Org. Chem.* **30**, 1693 (1965).

⁴⁹ H. KOBAYASHI and S. AKIYOSHI, *Bull. Chem. Soc. Japan* **35**, 1044 (1962), *ibid.* **36**, 823 (1963).

⁵⁰ W. W. REID, *Biochem. J.* **100**, 13P (1966).

⁵¹ J. A. GILES and J. N. SCHUMACHER, *Tetrahedron* **14**, 246 (1961).

pointed out previously,⁴⁶ the lack of head-to-tail arrangement in this compound. It should also be noted that cembrene⁴⁸ (33, thunbergene)⁴⁹ the parent hydrocarbon and expected precursor of the above mentioned compounds (29, 34–39) has only been isolated from tissue slices of *N. tabacum after inhibition of sterol synthesis by addition of tris(2-diethylaminoethyl)phosphate and β -diethylaminoethyl diphenylpropylacetate chloride.⁵⁰ A further indication that these compounds are formed by non-enzymatic reactions is the co-occurrence of diastereomers, which is rarely observed for diterpenoids from other natural sources and which in the case of tobacco is further illustrated by the presence of α - and β -levantenolides.⁵¹*



SCHEME 1 POSSIBLE RELATIONSHIP BETWEEN SOME TOBACCO ISOPRENOIDS
The formulae have been drawn in the above manner to account for the known stereochemistry of double bonds in 34, all-*trans*, but do not imply any other configurational assignment.

Four of the compounds given in Table 1 (1, 3, 26, 32) and β -ionone (40), encountered in a new larger batch of Greek tobacco presently being investigated in this laboratory, possess carbon skeletons identical to those encountered as terminal parts of many carotenoids, e.g. β -carotene. It has recently been demonstrated that β -ionone (40) and dihydroactinidiolide (32) are formed from β -carotene on photooxidation.⁵² Moreover, recent work on photooxidation of carotenoids and related compounds has shown that, in addition to the aforementioned oxidative cleavage of the 9,10-bond, similar fracture of the 11,12-bond and oxygenation of the 5,6 and 8 positions are encountered reactions.^{53–55} It seems conceivable therefore that the compounds β -cyclocitral, safranal, damascenone, dihydroactinidiolide, are formed in this manner. It is of interest that α - and β -carotene, lutein, zeaxanthin, cryptoxanthin, flavoxanthin and violaxanthin have been shown to be present in tobacco, and that many of these are adequate precursors for the compounds under consideration.⁵⁶

The acyclic nor-isoprenoids, which in contrast to solanone (29) display a regular arrangement of isoprene units, can be expected to be derived in a similar manner from higher non-

⁵² S. ISOE, S. B. HYEON and T. SAKAN, *Tetrahedron Letters* 279 (1969)

⁵³ M. MOUSSERON-CANET, J.-P. DALLE and J.-C. MANI, *Tetrahedron Letters* 6037 (1968)

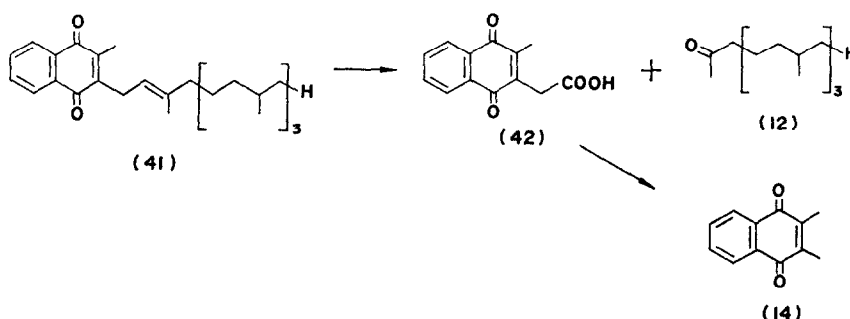
⁵⁴ C. S. FOOTE and M. BRENNER, *Tetrahedron Letters* 6041 (1968)

⁵⁵ R. S. BURDEN and H. F. TAYLOR, *Tetrahedron Letters* 4071 (1970)

⁵⁶ H. E. WRIGHT, JR., W. W. BURTON and R. C. BERRY, JR., *Arch. Biochem. Biophys.* **82**, 107 (1959).

cyclic isoprenoids Possible precursors so far encountered in tobacco for 6-methyl-5-hepten-2-one (31), geranylacetone (30), and farnesylacetone (27) are phytoene, phytofluene, solanesol and solanesyl derivatives, hexahydrofarnesylacetone (12) could similarly be derived from phytyl derivatives (*vide infra*)^{3,56} 6-Methyl-2-heptanone (28), 6-methyl-3,5-heptadien-2-one (4) and ψ -ionone (6) may arise from other isoprenoids so far not detected in tobacco

The occurrence of 2,3,6-trimethyl-1,4-naphthoquinone (13), which has previously been isolated from tobacco smoke,³⁰ and the corresponding 2,3-dimethyl derivative (14), is of interest in this context It seems conceivable that the latter arises from vitamin K₁ (41), a known tobacco constituent,⁵⁷ by oxidative cleavage of the phytyl double bond and subsequent decarboxylation of the resulting β,γ -unsaturated acid (42), see Scheme 2 The postulated formation of nor-solanesene,⁵⁸ recently isolated from the same Greek tobacco, provides an example of a similar route, also accomplished chemically, which involves oxidation of solanesol and decarboxylation of the resulting α,β -unsaturated acid It seems probable



SCHEME 2 POSSIBLE WAY OF FORMATION OF 2,3-DIMETHYL-1,4-NAPHTHOQUINONE (14) AND HEXA-HYDROFARNESYLACETONE (12) IN TOBACCO

that 2,3,6-trimethyl-1,4-naphthoquinone, readily distinguished from the corresponding 2,3,5-trimethyl derivative by comparison of mass spectra and retention times, may arise in a similar manner from compounds not yet detected in tobacco, e g 2,3-dimethyl-6-phytyl- or 2,6-dimethyl-3-phytyl-1,4-naphthoquinone Moreover, these reactions would, as mentioned above, also account for the formation of hexahydrofarnesylacetone (12) In view of the present findings, the earlier proposal³⁰ that 2,3,6-trimethyl-1,4-naphthoquinone may be formed in the smoking process from vitamin K₁ by thermal cleavage of the 1',2'-bond and subsequent methylation of C(6) in the resulting radical seems less satisfactory than simple transfer from the tobacco to the smoke

EXPERIMENTAL

Materials and methods The solvents, silica gel and drying agents used in the present study were purified as described earlier.² NMR spectra were recorded in CDCl₃ and IR spectra in CCl₄. GC-MS was carried out on a modified LKB 9000 instrument operated at 70 eV, in which the original gas chromatograph had been replaced by a home made unit equipped with a capillary injector, a device for introduction of make-up gas, a splitter and a flame ionization detector (FID). The signal for the FID could be detected a few sec prior to that of the total ion current sensor of the mass spectrometer due to different travelling time after the splitter.⁵⁹

⁵⁷ R. L. ROWLAND and J. A. GILES, *Tob. Sci.* **4**, 29 (1960)

⁵⁸ C. R. ENZELL, B. KIMLAND and L.-E. GUNNARSSON, *Tetrahedron Letters* 1983 (1971)

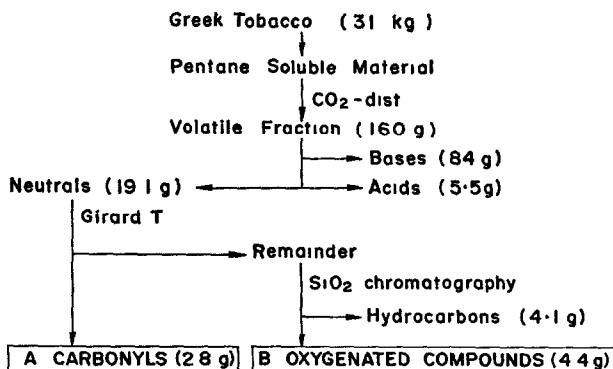
⁵⁹ J. ROERADE and C. R. ENZELL, *Acta Chem. Scand.* **22**, 2380 (1969)

The capillary columns used in the present study were made by coating stainless steel tubes (50 m \times 0.5 mm or 50 m \times 0.25 mm, Handy and Harman grade 316-S) with Apiezon L (Associated Electrical Industries, England) dissolved in hexane (10%), using the dynamic method⁶⁰

Analytical GLC was performed on a Varian 1700 instrument equipped with a capillary injector and a flame ionization detector. Preparative GLC was performed on columns (2 m \times 3.2 mm) packed with 3.5% OV 17 or 5% Carbowax on Chromosorb G using the same type of instrument but now equipped with an ordinary injector and an outlet splitter dividing the effluent between the FID and heated outlet tube in a 1:10 ratio. Fractions were collected in U-shaped teflon tubes (150 mm \times 2 mm) having one end inserted in the heated outlet tube, the center kept at about -70° and the other end equipped with an electrostatic precipitator⁶¹

Preparation of some reference compounds Hexahydrofarnesylacetone was prepared by hydrogenation of farnesylacetone⁶² (161 mg) in ethanol (3 ml) for 2 hr at room temp and atmos pressure using 10% Pd-C (20 mg) as catalyst. The reaction mixture was filtered, diluted with H₂O (30 ml) and subsequently extracted with pentane. Removal of the solvent left a colourless liquid (142 mg) which appeared to be homogeneous when gas chromatographed on a capillary column. Pure 6-methyl-2-heptanone (130 mg) was obtained in the same way by catalytic hydrogenation of 6-methyl-3,5-heptadien-2-one⁶³ (145 mg) using the same amounts of solvent and catalyst. 2-Acetylpyrrole was made according to the method of Oddo⁶⁴

Isolation The isolation of the fractions discussed in this paper has been described in detail previously.² Sun-cured Greek tobacco, *Nicotiana tabacum*, grown in Serres 1968, was extracted with pentane and ether. The pentane soluble material of the extracts was distilled *in vacuo* using CO₂ as carrier⁶⁵ as indicated in Scheme 3 (the weights indicated are not accurate due to the fact that the solvent has not been completely removed to avoid loss of very volatile material)



SCHEME 3 ISOLATION AND SEPARATION OF THE FRACTIONS INVESTIGATED

The distillate was extracted consecutively with aq H₂SO₄ and NaOH to eliminate acidic and basic material and give a neutral fraction. The neutral carbonyl containing compounds were partially separated via their water soluble Girard T salts⁶⁶ giving fraction A. The remaining neutral fraction was separated on silica gel into hydrocarbons and a fraction containing the more polar compounds, fraction B. Fractions A and B were rechromatographed separately on silica gel columns using pentane-ether mixtures as eluents to give seven (115, 73, 322, 256, 153, 196, 126 mg) and eight (412, 244, 177, 122, 228, 547, 895, 764 mg) fractions respectively. Each fraction was studied by GLC and by GC-MS. Most of the fractions were found to be complex and usually contained from 50 to 200 components. A limited number of the components could be isolated by preparative GLC (22 and 23) or by column chromatography on AgNO₃-impregnated silica gel⁶⁷ (12, 16, 27, 29, 30 and 32), see Table 1.

⁶⁰ G. DIJKSTRA and J. DE GOEY, in *Gas Chromatography* (edited by D. H. DESTY), p. 56, Butterworths, London (1958).

⁶¹ P. KRATZ, M. JACOBS and B. M. MITZNER, *Analyst* **84**, 671 (1959).

⁶² S. OSMAN and J. BARSON, *Tob. Sci.* **10**, 85 (1966).

⁶³ J. D. SURMATIS and A. OFNER, *J. Org. Chem.* **28**, 2735 (1963).

⁶⁴ B. ODDO, *Chem. Ber.* **43**, 1012 (1910).

⁶⁵ C. R. ENZELL, B. KIMLAND and A. ROSENGREN, *Acta Chem. Scand.* **24**, 1462 (1970).

⁶⁶ A. GIRARD and G. SANDULESCO, *Helv. Chim. Acta* **19**, 1095 (1936).

⁶⁷ T. NORIN and L. WESTFELT, *Acta Chem. Scand.* **17**, 1828 (1963).

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Key Word Index—*Nicotiana tabacum*, solanaceae, tobacco, terpenoids, norisoprenoids